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GENERALIZED URTICARIA AS A VASCULITIC MANIFESTATION IN A PATIENT WITH SARS-CoV-2 INFECTION: A CASE REPORT IN COLOMBIA

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KEYWORDS

SARS-CoV2

COVID-19

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ABSTRACT

Skin manifestations have been reported in up to 20% of cases of SARS-CoV-2 infection, including morbilliform rash (22%), pernio-like acral lesions (18%), urticaria (16%), and macular erythema (13%). It is believed that in the case of SARS-CoV-2 infection, the mechanism involved is an inflammatory response that generates immune dysregulation, vascular congestion, vasculitis, vascular thrombosis, or neoangiogenesis. This case study, present the case of a patient with no previous history of urticarial reactions, autoimmune diseases, or exposure to medications who develops generalized urticaria lasting more than 24 hours and who was diagnosed with SARS-CoV-2 infection by RT-PCR with a nasopharyngeal swab. We suggest in this patient vasculitic urticaria as a manifestation of SARS-CoV-2 infection.

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1 Introduction

The clinical spectrum of the coronavirus disease 2019 (COVID-19) (Dhama et al., 2020), caused by the severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2), is still complex (Yuki et al., 2020). This emerging disease, first reported in Wuhan, China (Lu et al., 2020), has extended globally (WHO, 2020) and still produces a vast number of clinical consequences (Ortiz-Maritnez et al., 2020).

Coronaviruses are enveloped, positive-sense, single-stranded RNA viruses that enter the cell through fusion with the receptor for angiotensin-converting enzyme 2 (ACE2), a well-known receptor for SARS-CoV, which is expressed in the cell membrane of numerous organs such as the heart, lung, ileum, central nervous system, kidney, and bladder (Hamming et al., 2004; Zhou et al., 2020). It is suggested that this interaction generates a down regulation of the expression of ACE2, an increase of angiotensin II due to unopposed accumulation, and activation of the reninangiotensin-aldosterone system (RAAS), which altogether play a role in the development of acute respiratory distress syndrome (ARDS) (Vaduganathan et al., 2020; Zhang et al., 2020a). Furthermore, it contributes to generating endothelial dysfunction, leading to vasoconstriction, increased vascular permeability, and abnormal remodeling of myocardial cells (Vaduganathan et al., 2020).

Transmission occurs mainly via respiratory droplets from direct person-to-person contact and, to a lesser degree, via fomites and aerosols (Wiersinga et al., 2020), with a high rate of spread (R₀ of 2.24-3.58) reported during the early phase of the outbreak (Chen, 2020; Zhao et al., 2020a). The majority of patients with SARS-CoV-2 pneumonia have a good prognosis, and the severity depends primarily on characteristics such as age, comorbidities, and the individual's immune condition (Wang et al., 2020). The gold standard for diagnosis is via viral RNA detection by reverse transcription-polymerase chain reaction (RT-PCR) (Corman et al., 2020; Loeffelholz et al., 2020; Sheikhzadeh et al., 2020). Skin manifestations have been reported in up to 20.4% of cases of SARS-CoV-2 infection (Recalcati, 2020), with an average onset of 9.92 days following the initiation of systemic symptoms (Zou et al., 2020; Zhao et al., 2020b). The most common manifestations include morbilliform rash (22%), pernio-like acral lesions (18%), urticaria (16%), and macular erythema (13%) (Freeman et al., 2020).

In SARS-CoV-2 infection, the suggested mechanism involves an inflammatory response that generates immune dysregulation, vascular congestion, vasculitis, vascular thrombosis, or neoangiogenesis. This response can affect the lungs, heart, liver, and kidney; leading to an increase in vascular permeability mediated by mast cell degranulation (Arias-Santiago et al., 2009)

and by basophil production of vasoactive amines; and complement activation mediated by circulating immune complexes (Leiste et al., 2008).

2 Case Report

A 34-year-old woman with sudden onset of generalized urticaria was predominantly localized to the lower extremities, with subsequent asymmetric, erythematous, round papules (Figure 1); first consulted to the outpatient clinic and was initially managed with antihistamines without clinical improvement. Two days later, she presents to the emergency department, denying any febrile episode since the onset of symptoms, with normal pulse oximeter readings and no respiratory symptoms. During the initial evaluation, a clinical presentation compatible with leukocytoclastic vasculitis was considered. An immunological profile with routine lab tests was requested, which show normal blood count levels, preserved kidney and liver function, CRP of 6.8 mg/L, negative antinuclear antibodies, complement, and IgE within normal limits. Management with corticosteroids is initiated, presenting a slight improvement of the lesions with the pruritus's persistence. Four days later, she starts presenting with dyspnea on exertion, requiring a new visit to the emergency department, where blood tests taken at the moment revealed neutrophilic leukocytosis without any other significant finding. The patient remained afebrile, with adequate oxygen saturation (95%) on a FiO₂ of 21%, and mild hypoxemia noted on arterial blood gas analysis.

Further testing showed ferritin of 1200 mcg/L and D-dimer of 6880 mcg/L. Given the presence of hypoxemia, a high-resolution computed tomography (HRCT) of the chest was ordered, which showed a discrete random interstitial compromise with an associated solitary lung nodule of indeterminate appearance localized to the right lower lobe; which, according to the official radiologic report, may correspond to a granuloma of probable residual origin and incipient paraseptal emphysema (Figure 2). Performing an active search for SARS-CoV2, a nasopharyngeal swab for RT-PCR was taken and reported as positive. Three days later, the pruritus and the cutaneous manifestations disappeared with no signs of persistent pulmonary compromise. On the seventh day, follow-up laboratory tests showed acute inflammatory markers within normal limits. On day fourteen, repeat RT-PCR testing for SARS-CoV-2 was reported as negative; the IgM rapid test reported positive with negative IgG. Laboratory findings are found in Table 1.

3 Discussion

The SARS-CoV-2 infection has become a relevant issue for public health at the moment. The associated cutaneous manifestations have been prevailing as a critical aspect to take into consideration in the clinical course of the disease, although the most frequent

Table 1 Laboratory findings in the patient with COVID-19

Blood lenkocyte count, 10°91. 37 - 10.1 1176 130 117 8.4	Table 1 Laboratory finding	gs in the patient w	ith COVID-19				
Blood leukocyte count, 10°9/L 3.7 · 10.1 11.76 13.0 11.7 8.4				DATE			
Blood leukocyte count, 10°9/L 3.7 · 10.1 11.76 13.0 11.7 8.4	Test		4/2020	5/2020	5/2020	5/2020	
Lymphoxyte, % 18.0 - 48.3 37.8 18.8 11.6 26.9		values	28/0	01/0	04/0	11/0	
Lymphocyte count, 10°9/L	Blood leukocyte count, 10^9/L	3.7 - 10.1	11.76	13.0	11.7	8.4	
Lymphocyte count, 10°9/L	Lymphocyte, %	18.0 - 48.3	37.8	18.8	11.6	26.9	
Neutrophil count, 10°9/L 1.63 - 6.96 6.26 9.2 7.2 4.7	Lymphocyte count, 10^9/L	1.09 - 2.99	4.45	2.46	2.36	2.27	
Neutrophili to lymphocyte ratio (NLR)	Neutrophil, %	39.3 - 73.7	53.3	70.7	78.2	56.5	
Monocyte count, 10°9/L 0.30 - 10.0 5.4 8.6 5.9 10.6	Neutrophil count, 10^9/L	1.63 - 6.96	6.26	9.2	7.2	4.7	
Monocyte count, 10%/L 0.3 - 0.9 0.64 1.13 0.69 0.89 Eosinophil, % 0.00 - 7.00 2.8 0.83 3.9 4.68 Eosinophil count, 10%/L 0 - 0.5 0.33 0.105 0.46 0.36	Neutrophil to lymphocyte ratio (NLR)	0.78 - 3.53		3.79		2.07	
Eosinophil, % 0.00 - 7.00 2.8 0.83 3.9 4.68	Monocyte, %	0.00 - 10.0	5.4	8.6	5.9	10.6	
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Basophil, % 0.00 - 1.00 0.5 0.9 0.4 1.20							
Basophil count, 10°9/L							
Erythrocyte count. 10^6/uL			0.5	0.9			
Hemoglobin, g/dL							
Hematocrit, % 37,7 - 53,7 40,6 45,1 42,3 43,9 Mean cell volume (MCV), fi. 79,0 - 101,0 83,4 86,7 88 86,3 Mean cell bemoglobin (MCH), pg 26,0 - 35,0 27,1 27,7 31,9 28 Mean corpuscular hemoglobin concentration (MCHC), g/dL 31,0 - 37,0 32,5 32,0 32 32,4 Red blood cell distribution width (RDW), µm 11,5 - 14,5 11,9 12,1 14 11,9 Platelet count, 10°9/L 150,450 318 317 313 382 Mean platelet volume, Fl 4,5 - 10 9,9 13,0 9,3 7,36 C-reactive protein level, mg/L 0 - 5 6,8 18 12 0,0 Ferritin (mcg/L) < 300 - 1200 620 215 D - dimer (mcg/L) < 500 - 6880 920 440 TSH (mUl/L) 0,46 - 4,68 2,5 - - - T4 (ng/dL) 0,89 - 1,76 0,92 - - - T3L (pg/dL) 1,8 - 4,2 4,27 - - - C3 (ng/dL) 1,8 - 4,2 4,27 - - - C4 (ng/dL) 2,41 - - - Eosinophil count 3% - - - Eosinophil count 3% - - - Antibodies Anti Rnp: < 20 5,39 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies Anti Sa (Ro): < 20 1,96 - - - Antibodies A							
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Mean cell hemoglobin (MCH), pg							
Mean corpuscular hemoglobin concentration (MCHC), g/dL 31.0 - 37.0 32.5 32.0 32 32.4 Red blood cell distribution width (RDW), µm 11.5 - 14.5 11.9 12.1 14 11.9 Platelet count, 10^9/L 150-450 318 317 313 382 Mean platelet volume, Fl 4.5 - 10 9.9 13.0 9.3 7.36 C-reactive protein level, mg/L 0 - 5 6.8 18 12 0.0 Ferritin (mcg/L) < 500 - 6880 920 240 D-dimer (mcg/L) < 500 - 6880 920 440 TSH (mU/L) 0.46 - 4.68 2.5 - - - T3L (pg/dL) 1.8 - 4.2 4.27 - - - C3 (ng/dL) 117 - - - C4 (ng/dL) 24.1 - - - Eosinophil count 33% - - - E-Inmunoglobulin <150 IU/ml 50.4 Antibodies Antinuclear Non Reactive - - - Antibodies Antin Smp: < 20 5.39 - - - Antibodies Antin Smp: < 20 4.41 - - - Antibodies Anti Sas (Ro): < 20 1.053 - - Antibodies Antin Sab (La): < 20 10.53 - - C-ANCAS PR3 > 1/20 Negative - - - Arterial-blood gases P-ANCAS MPO > 1/20 Negative - - - Arterial-blood gases P-ANCAS MPO > 1/20 Negative - - - Arterial-blood gases - - - - Arterial-blood gases - - - - - - Arterial-blood gases - - - - - - Arterial-blood gases - - - - - - - Arterial-blood gases - - - - - - - Arterial-blood gases - - - - - - - Arterial-blood gases - - - - - - - - Arterial-blood gases - - - - - - - - - Arterial-blood gases - - - - - - - - - Arterial-blood gases - - - - - - - - -							
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P-ANCAS MPO > 1/20 Negative - - - Arterial-blood gases - 7.36 - 7.44 - 7.39 7.37 7.43 pCO2 (mmHg) 33 - 40 - 41 41 34.6 pO2 (mmHg) 70 - 80 - 66.3 70.2 83 HCO3 (mmol/L) 21 - 27 - 21 22 23 BEb Base Excess (mmol/L) -3 to +3 - 0 0 0 SO2 % - 97 96 98 Temperature (°C) - 37.4 36.7 37.3 FIO2 % - 21 21 21 21 RT - PCR SARS-CoV-2 Positive Negative SARS - CoV2 (COVID - 19) Antibodies IgM Negative Positive				-	-	_	
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						Negative	
		<u> </u>	<u> </u>			Positive	

Bold, altered values



Figure 1 Cutaneous findings in the patient

initial symptoms are fever, fatigue, cough, dyspnea, myalgia, headache, and diarrhea; and to a lesser extent, nasal congestion, runny nose, and odynophagia. In severe cases, dyspnea and hypoxemia rapidly progress to ARDS, septic shock, refractory metabolic acidosis, and multi-organ dysfunction due to gastrointestinal complications, acute heart injury, acute kidney failure coagulopathy (Rossi et al., 2003; Huang et al., 2020). Mortality is mainly dependent on ARDS and its sequelae, along with an exaggerated inflammatory response called a cytokine storm (Zhang et al., 2020b).

Skin manifestations have been reported in up to 20.4% of cases of SARS-CoV-2 infection (Recalcati, 2020), with an average onset of 9.92 days posterior to the initiation of systemic symptoms (Zhao et al., 2020b). Among these manifestations, the most common include morbilliform rash (22%), pernio-like acral lesions (18%), urticaria (16%), and macular erythema (13%). Less frequently,

vesicular eruption (11%), papulosquamous eruption (9.9%), and retiform purpura (6.4%) (Freeman et al., 2020). Regarding the pathophysiology of SARS-CoV-2 infection, the mechanism involved remains unclear but suggested evidence points toward a possible immune dysregulation, vascular congestion, vasculitis, vascular thrombosis, or neoangiogenesis. All of which have their impact on the lungs, heart, liver, and kidney.

Multiple mechanisms have been proposed as triggers for this type 1 hypersensitivity reaction, most of which lead to increased vascular permeability. Some authors have suggested a cross-reaction between IgM or IgG antibodies, secondary to the infection, with IgE localized on mast cells' surface, which promotes mast cell degranulation (Arias-Santiago et al., 2009). In contrast, other groups defend that circulating immune complexes stimulate vasoactive amines by basophils and activate complement (Leiste et al., 2008). As a response to viral infection, cells of the



Figure 2 High-resolution computed tomography (HRCT) of the chest

immune system, including mast cells; found in the submucosa of the respiratory tract and the nasal cavity, release a series of inflammatory markers, including protease and cytokines, such as IL-1, IL-6, or IL-33; and histamine (Kritas et al., 2020). Such histamine release triggers the appearance of hives and angioedema. Vasculitic changes in SARS-CoV-2 infections are mainly related to the lymphocyte response and exhibit a few histopathological characteristics, such as dermal edema, vacuolar alteration of the basal layer, keratinocyte necrosis. A mixed infiltrate with a

predominant perivascular and peri-eccrine distribution in the papillary (superficial) and reticular (deep) layers of the dermis and the hypodermis (Kolivras et al., 2020; Recalcati et al., 2020; Romaní et al., 2020;); however, these vasculitic changes may also be related to a pauci-inflammatory thrombogenic vasculopathy in patients with severe SARS-CoV-2 infection due to activation of the complement pathways and an associated procoagulant state (Magro et al., 2020; Zhang et al., 2020c).

We present a patient with no previous history of urticarial reactions, autoimmune diseases, or exposure to medications that develops a widespread urticaria lasting more than 24 hours SARS-CoV-2 infection is diagnosed employing a nasopharyngeal swab for RT-PCR. Vasculitic urticaria is established as an associated manifestation of SARS-CoV-2 in this patient; therefore, although not frequently documented, we highlight the importance of dermatological findings as possible clinical pictures of the disease; and consequently, we emphasize the need for adequate precautions when evaluating patients who present to the clinical setting with cutaneous symptoms during the pandemic. That could have a significant impact on the prevention of the spread of the infection. Only time will elucidate the behavior and implications of these findings.

Author contributions

All the authors substantially contributed to the conception, compilation of data, checking and approving the final version of the manuscript, and agree to be accountable for its contents.

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Consent for publication

Written consent from the patient was obtained for publication.

Competing interests

The authors declare that they have no competing interests.

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